

# 2x PCR Master mix Solution ( i-StarMAX™ II )

The Best Choice of 2X Master for high-fidelity amplification of PCR targets up to 20 Kb.

**RUO** Research Use Only    **REF** 25174     $\Sigma$  100    2°C 8°C

## DESCRIPTION

iNtRON's Maxime PCR PreMix Kit has not only various kinds of PreMix Kit according to experience purpose, but also a 2X Master mix solution. **2x PCR Master mix Solution (i-StarMAX™ II)** is made from iNtRON's i-StarMAX™ II DNA Polymerase. Taq DNA Polymerase is the most common PCR enzyme for amplifying up to 10 kb IDNA templates and up to 3 kb genomic templates. However, it is not able to amplify even longer DNA fragments. To overcome this constraint, variable PCR systems have been developed by various company, which are containing Taq DNA Polymerase and thermostable DNA Polymerase with proofreading activity. i-StarMAX™ II DNA Polymerase cannot be useful tool only in amplification of short and long fragments but also problematic template/primer systems with hot-start function. High yields of PCR product can be achieved using extension times as short as from 30 seconds to 1 minute per kb per cycle with the i-StarMAX™ II DNA Polymerase. The i-StarMAX™ II DNA Polymerase is recommended for relatively rapid, high-fidelity amplification of PCR targets up to 20 kb when proofreading DNA polymerase alone requires too long an extension time or yields are insufficient.

**2x PCR Master mix Solution (i-StarMAX™ II)** is the product what is mixed every component. i-StarMAX™ II DNA Polymerase, dNTP mixture, reaction buffer, and so on in one tube. This is the product that can get the best result with the most convenience system. The first reason is that it has every components for PCR, so we can do PCR just add a template DNA, primer set, and D.W.. The second reason is that it has Gel loading buffer to do electrophoresis, so we can do gel loading without any treatment. In addition, each batches are checked by a thorough Q.C., so its reappearance is high. It is suitable for various sample's experience by fast and simple using method.

## CHARACTERISTICS

- **High fidelity, specificity (hot-start function), and yields**
- **Increased yield of PCR amplification**  
Because the accuracy of the enzyme blend reduces the number of truncated amplification products formed.
- **Improved performance of long PCR**  
Because the reaction buffer and the enzyme blend are optimized for generation of certain length products
- **Flexibility**  
Available for various DNA template including cloned fragment, phage DNA, mammalian genomic DNA and etc.
- **Ready to use** : only template and primers are needed
- **Construct as various reaction volume**
- **Stable for 18 months at 4 °C**
- **Time-saving and cost-effective**

## KIT CONTENTS

Contents	Amount
2X PCR Master mix Solution (i-StarMAX™ II)	0.5 ml x 2 Vials
Instruction Manual	1 ea

Component	Amount
i-StarMAX™ II DNA Polymerase (5 U/μl)	2.5 U
dNTPs	2.5 mM each
PCR Reaction Buffer	1x
Gel Loading buffer	1x

## STORAGE

- Storage condition : Store the product at 2 ~ 8 °C after receiving.
- Expiration : **2X PCR Master mix Solution (i-StarMAX™ II)** can be stored for up to 18 months without showing any reduction in performance and quality under appropriate storage condition. The expiration date is labeled on the product box.

## APPLICATIONS

- General PCR
- Disease diagnosis
- SNP analysis
- PCR for difficult template or primer such as human genomic DNA
- Multiplex PCR
- TA cloning & Blunt Cloning etc

## PRODUCT WARRANTY AND SATISFACTION GUARANTEE

All products undergo the thorough quality control test and are warranted to perform as described when used correctly. Immediately any problems should be reported. At iNtRON we pride ourselves on the quality and availability of our technical support. Our Technical Service Departments are staffed by experienced scientists with extensive practical and theoretical expertise in molecular biology and the use of iNtRON products. If you have any questions or experience any difficulties regarding the **2X PCR Master mix Solution (i-StarMAX™ II)** or iNtRON products in general, please do not hesitate to contact us. iNtRON customers are a major source of information regarding advanced or specialized uses of our products. This information is helpful to other scientists as well as to the researchers at iNtRON. We therefore encourage you to contact us if you have any suggestions about product performance or new applications and techniques. Satisfaction guarantee is conditional that the customer should provide full details of the problem to iNtRON within 60 days, and returning the product to iNtRON for examination.

## QUALITY CONTROL

In accordance with iNtRON's ISO-certified Total Quality Management System, each lot of **2X PCR Master mix Solution (i-StarMAX™ II)** is tested against predetermined specifications to ensure consistent product quality.

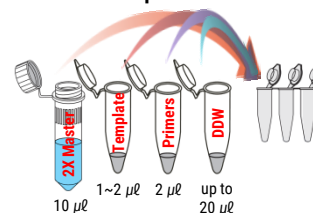
Contents	Quality Control
PCR Buffer, dNTP Mixture	Conductivity, pH, sterility, and performance in PCR are tested.
Distilled Water	Conductivity, pH, sterility, and performance in PCR are tested. Endonuclease, exonuclease, and RNase activities are tested.
2X PCR Master mix Solution (i-StarMAX™ II)	PCR reproducibility assay: The PCR reproducibility assay reactions are performed in using 3 batches.
Process Inspection	Accuracy of aliquot process was validated Appearance of Master mix solution (housing, sealing contamination)

## ADDITIONAL REQUIRED EQUIPMENT

- Distilled water
- Primers
- Pipettes and pipette tips (aerosol resistant)
- Thermal cycler
- Mineral oil (only if the thermal cycler does not have a heated lid)

## QUICK GUIDE

### 1. Mix the Components



### 2. Run the PCR Cycler



### 3. Electrophoresis of DNA



## PROTOCOL

**1. Dispense 10 µl of 2X PCR Master mix Solution (in case of total 20 µl PCR reaction) / 25 µl of 2X PCR Master mix Solution (in case of total 50 µl PCR reaction) into PCR tubes.**

**2. Add template DNA and gene specific primers into upper PCR tubes.**

**Note 1 :** Appropriate amounts of DNA template samples

- cDNA : 0.5-10% of first RT reaction volume
- Plasmid DNA : 10 pg-100 ng
- Genomic DNA : 0.1-1 µg for single copy

**Note 2 :** Appropriate amounts of primers

- Primer : 5-20 pmole/µl each (sense and anti-sense)

**3. Add distilled water into the tubes to make total volume 20 µl or 50 µl**

**Note:** This example serves as a guideline for PCR amplification. Optimal reaction conditions such as amount of template DNA and amount of primer, may vary and must be individually determined.

Example	PCR reaction mixture	20 µl Rxn	50 µl Rxn
	2X PCR Master mix Solution (i-StarMAX™ II)	10 µl	25 µl
	Template DNA	1 ~ 2 µl	1 ~ 2 µl
	Primer (F : 10 pmol/µl)	1 µl	1 µl
	Primer (R : 10 pmol/µl)	1 µl	1 µl
	Distilled Water	6 ~ 7 µl	21 ~ 22 µl
	<b>Total reaction volume</b>	<b>20 µl</b>	<b>50 µl</b>

**4. Mix the mixture well by pipetting or vortexing then spin down the mixture by brief centrifugation.**

**5. (Option) Add mineral oil.**

**Note:** This step is unnecessary when using a thermal cycler that employs a top heating (general methods) .

**6. Perform PCR of samples.**

**Note :** SUGGESTED CYCLING PARAMETERS

**\* Cycle program for fragments < 10kb**

	Temp.	Time	Cycle No.
Initial Denaturation	92 ~ 94°C	2 ~ 4min	1
Denaturation	94°C	15s - 1min	25-30
Annealing	45 ~ 65°C	15s - 1min	
Extension*	72°C	1min / 1 ~ 1.5kb	
Final extension	72°C 4°C	5 ~ 10min Hold	1

\*, Extension time for 30s-1min is sufficient for fragments < 1kb.

**\* Cycle program for fragments > 10kb**

	Temp.	Time	Cycle No.
Initial Denaturation	92 ~ 94°C	2 ~ 4min	1
Denaturation	94°C	15s - 1min	10
Annealing	45 ~ 65°C	15s - 1min	
Extension	72°C	1min / 1 ~ 1.5kb	
Denaturation	94°C	15s - 1min	15-20
Annealing	45 ~ 65°C	15s - 1min	
Extension	72°C	1min / 1 ~ 1.5kb + 20s / cycle	
Final extension	72°C 4°C	5 ~ 10min hold	1

**Note :** This CYCLING PARAMETERS serves as a guideline for PCR amplification. optimal reaction conditions such as PCR cycles, annealing temperature, extension temperature and incubation times, may vary and must be individually determined.

**7. Load samples on agarose gel without adding a loading-dye buffer and perform electrophoresis.**

**Note:** > 2% agarose (Agarose LE, iNtRON, Cat. No. 32034) gel is recommended.

## TROUBLESHOOTING GUIDE

### Symptoms & Possible Causes

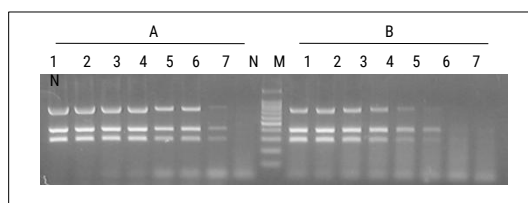
### Comments & Suggestions

Little or no product	•Repeat the PCR. Check the concentrations and storage conditions of the kit, primers and template.
A. Pipetting error or missing reagent	•Repeat the PCR with different primer concentrations from 0.1-0.5 µM of each primer (in 0.1 µM increments).
B. Primer concentration is not optimal or primers degraded	•Check the concentration, storage conditions, and quality of the starting template. If necessary, make new serial dilutions of template nucleic acid from stock solutions. Repeat the PCR using the new dilutions.
C. Problems with starting template	•Increase the number of cycles in increments of 5 cycles.
D. Insufficient number of cycles	•Check whether PCR was started with an initial denaturation step at 95°C for 5 min.
E. Hot start function is not activated	•Decrease annealing temperature by 2°C increments.
F. Incorrect annealing temperature or time	•Annealing time should be between 1 - 2 m
G. Incorrect denaturation temperature or time	Adjust the time in increments of 5 s.
H. Extension time too short	•Increase the extension time by increments of 30 s.
I. Primer design is not optimal	•Review primer design.
J. cDNA concentration	•For RT-PCR, take into consideration the efficiency of reverse transcriptase reaction which averages 10-30%. As RT reaction Mix is known to be a PCR inhibitor. The added volume of the cDNA should not exceed 10% of the final PCR volume.
Dimer or Product bands are smeared	
A. Primer concentration is not optimal or primers degraded	•Repeat the PCR with different primer concentrations from 0.1-0.5 µM each.
B. Primer design is not optimal	•Primer (in 0.1 µM increments). Review primer design.
C. Cycle number is too high	•Reduce the cycle number in increments of three cycles.
D. Quality of template DNA is too low	•Always use high-quality, purified DNA templates.
E. Carryover contamination	•Dispose of reagents, make fresh reagents, then repeat the PCR.

## EXPERIMENT INFORMATIONS

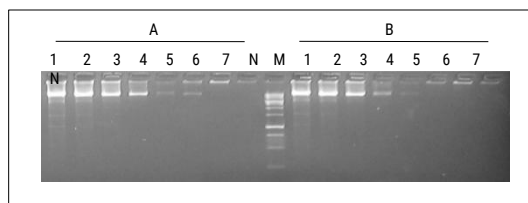
### ❖ Comparison test for sensitivity

After serial dilution of E.coli genomic DNA template, multiplex PCR for *fyuA*(780 bp), *tsh*(420 bp), *Irp2*(280 bp) is performed. Comparing to competitor's product, 2X PCR Master mix Solution(i-StarMAX™ II) is higher than competitor's product.



**A,** 2x PCR Master mix Solution(i-StarMAX™ II); **B,** Competitor  
**Lane M,** 100 bp DNA Marker; **lane N,** Negative control; **lane 1,** 100 ng gDNA; **lane 2,** 50 ng gDNA; **lane 3,** 25 ng gDNA; **lane 4,** 12.5 ng gDNA; **lane 5,** 6.25 ng gDNA; **lane 6,** 3.125 ng gDNA; **lane 7,** 1.5625 ng gDNA

The competitor's product of the same type was used for long PCR of 20 Kb fragment which was lambda DNA template serially diluted. Data shows 2X PCR Master mix Solution(i-StarMAX™ II) has higher sensitivity than competitor's product.



**A,** 2x PCR Master mix Solution(i-StarMAX™ II); **B,** Competitor  
**Lane M,** 1 Kb DNA Marker; **lane N,** Negative control; **lane 1,** 50 ng DNA; **lane 2,** 25 ng DNA; **lane 3,** 12.5 ng DNA; **lane 4,** 6.25 ng DNA; **lane 5,** 3.125 ng DNA; **lane 6,** 1.56 ng DNA; **lane 7,** 0.753 ng DNA